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(54) Title: POLYMERIC, FIBER MATRIX DELIVERY SYSTEMS FOR BIOACTIVE COMPOUNDS

(57) Abstract: Multifunctional systems for delivery of bioactive compounds incorporated within or between polymeric fibers in a matrix are provided. Also provided are methods of delivering bioactive compounds via implementation, coating and/or wrapping of these systems and methods for modulating the rate of release of bioactive compounds from these delivery systems.

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**POLYMERIC, FIBER MATRIX DELIVERY SYSTEMS  
FOR BIOACTIVE COMPOUNDS**

**Field of the Invention**

The present invention relates to delivery systems  
5 comprising polymeric fiber matrices, film coatings or  
braided/woven structures for the controlled release of  
bioactive compounds. The delivery systems of the present  
invention may be comprised of either biodegradable or  
nondegrading polymeric fibers. In one embodiment, these  
10 fibers have submicron and/or micron diameters. Bioactive  
compounds are included in the delivery system either by  
suspending the compound particles or dissolving the compound  
in the polymer solution used to produce the fibers.

**Background of the Invention**

15 A number of polymer matrices for use in the controlled  
release and/or delivery of bioactive compounds, and for  
particular drugs, have been described.

U.S. Patent 3,991,766 describes a medicament repository  
consisting of a surgical element in the form of tubes, sheets,  
20 sponges, gauzes or prosthetic devices of polyglycolic acid  
having incorporated therein an effective amount of a  
medicament.

U.S. Patent 4,655,777 describes a method for producing  
a biodegradable prosthesis or implant by encasing an effective  
25 amount of fibers of calcium phosphate or calcium aluminate in  
a matrix of polymer selected from the group consisting of  
polyglycolide, poly(DL-lactide), poly(L-lactide),  
polycaprolactone,, polydioxanone, polyesteramides,  
copolyoxalates, polycarbonates, poly(glutamic-co-leucine) and

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blends, copolymers and terpolymers thereof to form a composite.

U.S. Patent 4,818,542 discloses a method for preparing a spherical microporous polymeric network with interconnecting  
5 channels having a drug distributed within the channels.

U.S. Patent 5,128,170 discloses a medical device and methods for manufacturing medical devices with a highly biocompatible surface wherein hydrophillic polymer is bonded onto the surface of the medical device covalently through a  
10 nitrogen atom.

U.S. Patent 5,545,409 discloses a composition and method for controlled release of water-soluble proteins comprising a surface-eroding polymer matrix and water-soluble bioactive growth factors.

15 U.S. Patent 5,769,830 discloses synthetic, biocompatible, biodegradable polymer fiber scaffolds for cell growth. Fibers are spaced apart by a distance of about 100 to 300 microns for diffusion and may comprise polyanhydrides, polyorthoesters, polyglycolic acid or polymethacrylate. The  
20 scaffolds may be coated with the materials such as agar, agarons, gelatin, gum arabic, basement membrane material, collagen type I, II, III, IV or V, fibronectin, laminin, glycosaminoglycans, and mixtures thereof.

U.S. Patent 5,898,040 discloses a polymeric article for  
25 use in drug delivery systems which comprises a polymeric substrate with a highly uniform microporous polymeric surface layer on at least part of the substrate.

Encapsulation of a bioactive compound within a polymer matrix has also been described. For example, WO 93/07861  
30 discloses polymer microspheres of 50 to 100 microns comprising a compound contained in a fixed oil within the polymer microsphere. U.S. Patent 5,969,020 discloses a foam precursor comprising a crystalline thermoplastic polymer and solid crystalline additive for use in preparation of drug delivery  
35 systems.

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Recently, it has been shown that polymer fibers of nanometer diameter can be electrospun from sulfuric acid into a coagulation bath (Reneker, D.H. and Chun, I. Nanotechnology 1996 7:216). In these studies more than 20 polymers including  
5 polyethylene oxide, nylon, polyimide, DNA, polyaramide and polyaniline were electrospun into electrically charged fibers which were then collected in sheets or other useful geometrical forms. Electrospinning techniques have also been applied to the production of high performance filters (Doshi,  
10 J. and Reneker, D.H. Journal of Electrostatics 1995 35:151; Gibson et al. AIChE Journal 1999 45:190) and for scaffolds in tissue engineering (Doshi, J. and Reneker, D.H. Journal of Electrostatics 1995 35:151; Ko et al. "The Dynamics of Cell-Fiber Architecture Interaction," Proceedings, Annual Meeting,  
15 Biomaterials Research Society, San Diego, CA, April 1998; and WO 99/18893). WO 99/18893 describes a method for preparing nanofibrils from both nondegrading and biodegradable polymers for use as tissue engineering scaffolds.

The present invention relates to delivery systems for  
20 the controlled release of bioactive compounds which comprise polymeric fibers and the bioactive compound.

#### Summary of the Invention

An object of the present invention is to provide a system for delivery of bioactive compounds comprising a  
25 bioactive compound incorporated within or between a polymeric fiber matrix or linear assembly, film coating or braided/woven structure.

Another object of the present invention is to provide a method for delivering a bioactive compound to a patient for  
30 controlled release of the bioactive compound in the patient. In one embodiment of this method of the present invention, the bioactive compound is incorporated into a polymeric fiber matrix or linear assembly or a braided or woven structure and implanted into the patient. In another embodiment, the

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bioactive compound is incorporated into a polymeric fiber film used to coat implants, tissue engineering scaffolds and other devices such as pumps and pacemakers which are then implanted into the patient. In yet another embodiment, the bioactive  
5 compound is incorporated into a polymeric fiber film used to wrap organs, tissues or vessels in a patient.

Another object of the present invention is to provide methods for modulating the rate of release of a bioactive compound from a delivery system for bioactive compounds  
10 comprising a bioactive compound incorporated within or between polymeric fibers. These methods include modulating loading of the bioactive compound incorporated with or between polymeric fiber, selecting polymers to produce the polymeric fibers which degrade at varying rates, varying polymeric  
15 concentration of the polymeric fibers and varying polymeric fiber diameter.

#### Detailed Description of the Invention

Electrospinning is a simple and low cost electrostatic self-assembly method capable of fabricating a large variety  
20 of long, meter-length, organic polymer fibers with micron or submicron diameters, in linear, 2-D and 3-D architecture. Electrospinning techniques have been available since the 1930's (U.S. Patent 1,975,504). In the electrospinning process, a high voltage electric field is generated between  
25 oppositely charged polymer fluid contained in a glass syringe with a capillary tip and a metallic collection screen. As the voltage is increased, the charged polymer solution is attracted to the screen. Once the voltage reaches a critical value, the charge overcomes the surface tension of the  
30 suspended polymer cone formed on the capillary tip of the syringe and a jet of ultrafine fibers is produced. As the charged fibers are splayed, the solvent quickly evaporates and the fibers are accumulated randomly on the surface of the collection screen. This results in a nonwoven mesh of nano

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and micron scale fibers. Varying the charge density (applied voltage), polymer solution concentration, solvent used, and the duration of electrospinning can control the fiber diameter and mesh thickness. Other electrospinning parameters which  
5 may be varied routinely to effect the fiber matrix properties include distance between the needle and collection plate, the angle of syringe with respect to the collection plate, and the applied voltage.

In the present invention, electrospinning is used to  
10 produce polymeric fiber matrices with the capability of releasing bioactive compounds in a controlled manner over a selected period of time. In one embodiment, the delivery system of the present invention is used to maintain delivery of a steady concentration of bioactive compound. In another  
15 embodiment, the delivery system is used in pulsed delivery of the bioactive compound wherein the compound is released in multiple phases in accordance with either rapid or slow degradation of the polymer fibers or diffusion of the bioactive compound from the polymer fibers. In yet another  
20 embodiment, the delivery system is used to obtain a delayed release of a bioactive compound. For example, the bioactive compound-containing fiber polymer matrix can be coated with a layer of nonwoven polymer fiber matrix with no bioactive compound. In this embodiment, different polymers with  
25 different degradation times can be used to obtain the desired time delays.

The delivery systems of the present invention can be used to deliver a single bioactive compound, more than one bioactive compound at the same time, or more than one  
30 bioactive compound in sequence. Thus, as used herein, the phrases "a bioactive compound" and "the bioactive compound", are meant to be inclusive of one or more bioactive compounds.

For purposes of the present invention by "fiber" it is meant to include fibrils ranging in diameter from submicron,  
35 i.e. approximately 1 to 100 nanometers ( $10^{-9}$  to  $10^{-7}$  meters) to

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micron, i.e. approximately 1-1000 micrometers. The bioactive compound is incorporated within the polymeric fibers either by suspension of compound particles or dissolution of the compound in the solvent used to dissolve the polymer prior to  
5 electrospinning of the polymeric fibers. For purposes of the present invention, by "incorporated within" it is meant to include embodiments wherein the bioactive compound is inside the fiber as well as embodiments wherein the bioactive compound is dispersed between the fibers. The polymeric  
10 fibers comprising the bioactive compound can be arranged as matrices, linear assemblies, or braided or woven structures. In addition, the fibers which release a bioactive compound can serve as film coatings for devices such as implants, tissue engineering scaffolds, pumps, pacemakers and other composites.

15 These fiber assemblies can be spun from any polymer which can be dissolved in a solvent. The solvent can be either organic or aqueous depending upon the selected polymer. Examples of polymers which can be used in production of the polymeric fibers of the present invention include, but are not  
20 limited to, nondegradable polymers such as polyethylenes, polyurethanes, and EVA, and biodegradable polymers such as poly(lactic acid-glycolic acid), poly(lactic acid), poly(glycolic acid), poly(glaxanone), poly(orthoesters), poly(pyrolic acid) and poly(phosphazenes).

25 Examples of bioactive compounds which can be incorporated into the polymeric fibers include any drug for which controlled release in a patient is desired. Some examples include, but are not limited to, steroids, antifungal agents, and anticancer agents. Other bioactive compounds of  
30 particular use in the present invention include tissue growth factors, angiogenesis factors, and anti-clotting factors.

If the bioactive compound is to reside within or inside the polymer fiber, selection of the polymer should be based upon the solubility of the bioactive compound within the  
35 polymer solution. Water soluble polymers such as polyethylene

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oxide can be used if the bioactive compound also dissolves in water. Alternatively, hydrophobic bioactive compounds which are soluble in organic solvent such as steroids can be dissolved in an organic solvent together with a hydrophobic polymer such as polylactic glycolic acid (PLGA).

If the bioactive compound is to reside between the polymer fibers, dissolution of the bioactive compound in the polymer solution is not required. Instead, the bioactive compound can be suspended in the polymer solution prior to electrospinning of the fibers.

In one embodiment of the present invention, the bioactive compound-containing fibers can be splayed directly onto devices such as implants, tissue engineering scaffolds, pumps and pacemakers as a film coating. For implants and tissue engineering scaffolds, examples of preferred bioactive compounds include tissue growth factors and angiogenesis factors. For pumps or pacemakers, the bioactive compound may comprise an anti-clotting factor. The coated device is then implanted into a patient wherein the bioactive compound or compounds are released upon degradation of or by diffusion from, or combinations thereof, the polymeric fiber film.

In another embodiment, a matrix or linear assembly of the bioactive compound-containing fibers is prepared. In this embodiment, the matrix or linear assembly of bioactive compound-containing fibers can be sandwiched between layers of polymer which contain no bioactive compound to decrease any burst effect and/or to obtain a delayed release. Alternatively, the matrix may comprise layers of fibers containing different bioactive compounds. The matrix or linear assembly is then implanted into a patient for controlled release of the bioactive compound as the polymeric fibers degrade or as the bioactive compound diffuses from the polymeric fibers. The time delay can be controlled by varying the choice of polymer used in the fibers, the concentration of polymer used in the fiber, the diameter of the polymeric



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fibers, and/or the amount of bioactive compound loaded in the fiber.

For purposes of the present invention, by "implanting" or "implanted" as used herein, it is meant to be inclusive of placement of the delivery systems of the present invention into a patient to achieve systemic delivery of the bioactive compound, as well as placement of the delivery system into a patient to achieve local delivery. For example, the delivery systems of the present invention may be placed on the wound of a patient to enhance healing via release of the bioactive compound. Delivery systems may also be placed on the surface or wrapped around an organ, tissue or vessel for delivery of the bioactive compound to the organ tissue or vessel.

In another embodiment of the present invention, a braided, knitted or woven structure of bioactive compound-containing fibers is prepared. These structures are prepared using an extension of the traditional 2-dimensional braiding technology in which fabric is constructed by the intertwining or orthogonal interlacing of yarns to form an integral structure through position displacement. A wide range of 3-dimensional structures comprising the bioactive compound-containing fibers can be fabricated in a circular or rectangular loom. In this embodiment, the structure may comprise only bioactive compound-containing fibers, bioactive compound-containing fibers sandwiched between polymeric fibers which contain no bioactive compound, or a mixtures of fibers containing different bioactive compounds. Like the matrix or linear assembly, this structure can be implanted into a patient for controlled release of the bioactive compound or compounds as the polymeric fibers degrade or as the bioactive compound diffuses from the polymeric fibers. Again, delivery rate of the bioactive compound can be controlled by varying the choice of polymer used in the fibers, the concentration of polymer used in the fiber, the diameter of the polymeric

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fibers, and/or the amount of bioactive compound loaded in the fiber.

Accordingly, the present invention also relates to methods for modulating the rate of release of a bioactive compound from a delivery system for bioactive compounds comprising a bioactive compound incorporated within or between polymeric fibers. By "modulate" or "modulating", it is meant that the rate or release of the bioactive compound incorporated within or between the polymeric fibers of the delivery system is increased or decreased. Methods for modulating the rate of release include increasing or decreasing loading of the bioactive compound incorporated within or between the polymeric fibers, selecting polymers to produce the polymeric fibers which degrade at varying rates, varying polymeric concentration of the polymeric fibers and/or varying diameter of the polymeric fibers. Varying one or more of these parameters can be performed routinely by those of skill in the art based upon teachings provided herein.

The ability of systems of the present invention to release a bioactive compound in a controlled manner was demonstrated using polymeric fiber matrices containing fluorescently labeled bovine serum albumin (FITC-BSA) dispersed between the fibers of the matrix. To construct the bioactive compound-loaded matrices, various concentrations of finely ground FITC-BSA were suspended in biodegradable polymer polylactic glycolic acid in 50:50 dimethyl formamide:tetrahydrofuran. Suspensions contained in a glass syringe with a capillary tip were electrospun into approximately 500 nm diameter fibers via an electrostatic based self-assembly process in which a high voltage electric field was generated between the oppositely charged polymer and a metallic collection screen. At a critical voltage the charge overcomes the surface tension of the deformed polymer drop at the needle tip, producing an ultrafine jet. The similarly charged fibers are splayed and during their passage

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to the screen, the solvent quickly evaporates so that dry fibers accumulate randomly on the screen forming a mesh matrix.

The material properties of this mesh matrix of bioactive compound-containing fibers were examined via standard electron microscopy and tensile testing. It was found that tensile strength and the release profiles were a function of protein loading.

*In vitro* release of the FITC-BSA into an infinite sink of 37°C phosphate buffered saline was also measured. This sink mimics *in vivo* conditions. While release in the first 24 hours after initiation was dominant, release to over 120 hours was observed with an increase in release at the point where the fibers started to breakdown.

The following nonlimiting examples are provided to further illustrate the present invention.

#### EXAMPLES

##### Example 1: Preparation of fiber matrix containing BSA-FITC

A 25% (w/v) solution of polylactic glycolic acid was prepared in a 50:50 mixture of dimethylformamide and tetrahydrofuran. A mixture of FITC-BSA and BSA in the ratio of 1:5 was added to the solution in order to obtain 2% protein loading. A syringe containing 5 ml of the polymer plus bioactive compound mixture was placed at an angle of 45°. The syringe was fitted with a 16G needle with the tip of the needle at a distance of 24 cm from the metallic collection screen. A piece of nonwoven mat was placed on the metallic screen. A voltage of 20 kV was applied between the collection screen and the needle tip which resulted in fibers being sprayed into a nonwoven matrix on the metallic screen. The spraying was complete in about 4 hours.

It was found that with this specific polymer solvent system, polymer concentrations lower than 25% resulted in fibers with beads of polymers. These beads were eliminated when the polymer concentration was increased to 25% or

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greater. However, as will be understood by the skilled artisan upon reading this disclosure, this concentration will vary for different polymer/solvent systems and different bioactive compounds.

#### 5 Example 2: *In vitro* Release of Protein

*In vitro* release of the FITC-BSA into an infinite sink of 37°C phosphate buffered saline was measured. Pre-weighed pieces from different regions of the mat were placed into scintillation vials and 10 ml of phosphate buffered saline  
10 were added and the capped vials were placed on a rotary shaker at 37°C. The buffer was exchanged at different points in time in order to mimic infinite sink conditions. The amount of protein released was measured in the form of fluorescence of the FITC-BSA on a spectrophotofluorometer at an excitation  
15 wavelength of 495 nm and an emission wavelength of 513 nm.

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What is Claimed is:

1. A system for delivery of bioactive compounds comprising a bioactive compound incorporated within or between polymeric fibers.
- 5 2. The system of claim 1 which is biodegradable.
3. The system of claim 1 which is nondegradable.
4. The system of claim 1 wherein the fibers are arranged as a matrix or linear assembly, a film coating on a device, or a braided or woven structure.
- 10 5. The system of claim 1 wherein particles of the bioactive compound are suspended in a polymer solution prior to electrospinning of the polymeric fibers so that the bioactive compound is incorporated between the polymeric fibers.
- 15 6. The system of claim 1 wherein the bioactive compound is dissolved into a polymer solution prior to electrospinning of the polymeric fibers so that the bioactive compound is incorporated within the polymeric fibers.
7. The system of claim 1 comprising more than one  
20 bioactive compound incorporated into a single or multiple layers of polymeric fibers for delivery of the bioactive compounds sequentially or in concert.
8. A method for delivering bioactive compounds to a patient comprising incorporating a bioactive compound into a  
25 polymeric fiber matrix or linear assembly or a braided or woven structure and implanting the polymer fiber matrix or linear assembly or braided or woven structure into the patient.

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9. The method of claim 8 further comprising coating a device with the polymeric fiber matrix or linear assembly or braided or nonwoven structure and implanting the coated device into the patient for delivery of the bioactive  
5 compounds.

10. The method of claim 9 wherein the device comprises a tissue engineering device and the bioactive compound enhances cell attachment and growth to the device.

11. The method of claim 8 wherein the polymeric fiber  
10 matrix or linear assembly or a braided or woven structure is implanted directly on a wound of the patient to deliver the bioactive compound to the wound of the patient.

12. The method of claim 8 wherein the polymeric fiber matrix or linear assembly or a braided or woven structure is  
15 implanted on the surface of an organ, tissue or vessel of the patient to deliver the bioactive compound to the organ, tissue or vessel of the patient.

13. The method of claim 12 wherein the polymeric fiber matrix or linear assembly or a braided or woven structure is  
20 wrapped around the surface of an organ, tissue or vessel of the patient.

14. A method for modulating rate of release of a bioactive compound from a delivery system for bioactive compounds comprising a bioactive compound incorporated within  
25 or between polymeric fibers, said method comprising modulating loading of the bioactive compound incorporated with or between polymeric fiber, selecting polymers to produce polymeric fibers which degrade at varying rates, varying diameter of the polymeric fibers, or varying polymeric concentration of the  
30 polymeric fibers.

## INTERNATIONAL SEARCH REPORT

International application No.

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<b>A. CLASSIFICATION OF SUBJECT MATTER</b> IPC(7) : A61F 13/00, 2/00; A61K 9/14 US CL : 424/426, 422, 423, 424, 425, 486 According to International Patent Classification (IPC) or to both national classification and IPC		
<b>B. FIELDS SEARCHED</b> Minimum documentation searched (classification system followed by classification symbols) U.S. : 424/426, 422, 423, 424, 425, 486  Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched  Electronic data base consulted during the international search (name of data base and, where practicable, search terms used) CAS ONLINE, EAST		
<b>C. DOCUMENTS CONSIDERED TO BE RELEVANT</b>		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 99/18893 A1 (DREXEL UNIVERSITY) 22 APRIL 1999 (22.04.1999), see whole document.	1-14
X	US 5,545,409 A (LAURENCIN et al) 13 AUGUST 1996 (13.08.1996), see whole document.	1-14
Y	US 4,655,777 A (DUNN et al) 7 APRIL 1987 (07.04.1987), see whole document.	1,3-14
Y	US 5,324,519 A (DUNN et al) 28 JUNE 1994 (28.06.1994), see whole document.	1,3-14
<input type="checkbox"/> Further documents are listed in the continuation of Box C. <input type="checkbox"/> See patent family annex.		
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